Overexpression of the Acidic C-Terminus Region of HvRAF from Barley Confers Enhanced Cadmium Tolerance in Yeast

Jinwook Jung and Minkyun Kim*

School of Agricultural Biotechnology and Center for Agricultural Biomaterials, Seoul National University, Seoul 151-921, Korea

The barley ERF-type transcription factor, HvRAF, has an acidic C-terminus domain that contains a hemopexin domain signature-like sequence. As an initial study to investigate the functioning of this domain, we cloned the partial *HvRAF* gene (*pHvRAF*) corresponding to the domain into a yeast/*E. coli* shuttle vector, pYES2. We then introduced the pYES2::*pHvRAF* recombinant plasmid into yeast (*Saccharomyces cerevisiae* strain INVSc1). Here, we report that yeast transformants harboring pYES2::*pHvRAF* show significant tolerance to cadmium stress.

Keywords: cadmium stress, ERF-type transcription factor, Hordeum vulgare, Saccharomyces cerevisiae

Cadmium is extremely toxic in natural environments. When accumulated in excess within plant tissues, it can inhibit enzyme activity, membrane transport, photosynthesis, lipid peroxidation, etc. (Keck, 1978; Clijstersw and van Assche, 1985; Somashekaraiah et al., 1992; Krupa et al., 1993). In yeasts and higher plants, several components are involved in the cellular mechanism for cadmium tolerance, including phytochelatins, metallothioneins, glutathione, and an ATPbinding cassette (ABC) transporter (Li et al., 1996; Zenk, 1996; Lu et al., 1997; Cobbett et al., 1998; Lee et al., 2004). As an enzymatically synthesized peptide, phytochelatin binds heavy metals with high affinity, being sequestered to the vacuole. Therefore, the genes required for its synthesis play an important role in detoxifying cadmium. This involvement of phytochelatin in cellular defenses against cadmium stress has been well studied using two heterologous systems -- yeast and Arabidopsis. For example, overexpression of phytochelatin synthase genes from Arabidopsis and tobacco suppresses the cadmium-sensitive yap1 and ycf1 mutations in yeast (Clemens et al., 1999; Ha et al., 1999; Vatamaniuk et al., 1999; Kim et al., 2005). The yeast YAP1 gene encodes a transcription factor required for the expression of YCF1, an ABC transporter gene responsible for vacuolar sequestration of the complexes of cadmium with glutathione in yeast (Li et al., 1996). In addition, Arabidopsis over-expressing YCF1 shows enhanced resistance to cadmium and lead (Song et al., 2003).

Transcription factors play crucial roles as the final regulatory components in signaling pathways for defense responses against diverse environmental stresses, including heavy metal contamination. A number of transcription factors identified in higher plants contain various domains, e.g., ERF, bZIP, and WRKY (Chen et al., 2002). Previously, we reported the cloning and characterization of *HvRAF* in barley (<u>Hordeum vulgare Root Abundant Factor</u>, Genbank no. DQ102383), which encodes an ERF-type transcription factor (Jung et al., 2007). Its overexpression not only activates a variety of stress-inducible genes, but also confers enhanced

*Corresponding author; fax +82-2-873-3112 e-mail mkkim3@plaza.snu.ac.kr pathogen resistance and salt tolerance in *Arabidopsis*. The open reading frame of this gene, AAZ14068, encodes a

Α	GAG	CTG	ATG	GAG	TTT	TTC	AAC	GTG	GAC	GCC	ATC	GTC	CAC	CTG	ACC	ACT	GCC	GTC	GCC	GCG	60
	E	ι	N	Ε	F	F	N	۷	D	A	I	۷	н	L	T	τ	A	۷	A	A	20
	CTA	CCG	CCT	GTC	ACG	GCG	AGC	ACC	TTC	GCC	GAC	ACG	ATG	CCC	AGG	GTC	GAC	GAG	GAC	TCT	120
	L	Ρ	Ρ	۷	ĩ	A	\$	T	F	A	D	T	M	Ρ	R	۷	D	ε	D	s	40
	TCT	GTG	GGG	AGC	GGC	GGC	GGC	GCC	ATG	CTG	GGG	TTC	GCC	GAC	GAG	CTT	GGG	TTC	GAT	CCG	180
	s	۷	G	s	G	G	G	A	M	L	G	F	A	D	ε	L	G	F	D	Ρ	60
	TTC	ATG	ATG	TTC	CAG	CTA	000	TGC	TCC	GAC	ATG	TAC	GAA	TCC	GCO	GAC	AGC	ATC	TTC	GCC	240
	F	M	M	F	L	ι	Ρ	C	s	D	M	Y	ε	s	A	D	s	1	F	A	80
	GGA	GAC	GCT	GTC	ATC	XCCG	GAT	GCC	CTC	AGC	GTG	GAC	AGT	GGC	ATG	GAC	GCC	GTC	AGO	CTC	300
	G	D	A	۷	I	٩	D	A	L	s	۷	D	s	G	×	D	A	۷	s	ι	100
	TGO	AGC	TTC	GAC	GAG	TTC	ccc	ATG	GAC	AGC	GCC	ATT	TTC	TGA	CGC	111	ccc	TGT	GAT	GCA	360
	W	S	F	D	E	F	Ρ	M	D	s	A	I	F	•							
	ста	CAC	TCT	GTT	GGT	TGT	AAG	i AA	сто	CAC	CTG	GCC	TCT	ACG	TAG	TTC	CTT	GTA	AA 1	GCC	420
	CGC	GCA	CAG	AAC	CTT	GCT	CAG	ACC	AGA	TTC	TGT	TTC	TTG	GCC	AGG	AAC	GAA	AGG	AAG	GGT	480
	TGC	TGC	CGA	TGC	ATG	ATT	GCT	TCC	TCG	ATG	AAC	GCA	GAT	TCG		TGT	ATT	CTA	CTG	TTT	540
	GAG	ITT	CTT	GTT	CGI	CAC	ACA	CTG	TAC	CAA	ACT	GTA	TTG	TAC	CCT	ATC	ATA	ATT	TCT	GCT	600
	CGG	TAC	CTC	TCT	GTA	CTG	CTG	GTA	CCA	AAC	TGT	ATT	GTA	CTC	TGT	CAT	GAT	CTG	TAC	TAG	660
	TCT	TTG	GTA	CTG	CTO	GTO	AAT	AGT	100	ACG	AAT	TGA	TCA	AAA		AAA		AAA		AAA	720
	AAA			A A A	**																

в

	207 204 172	-GLDDVSLWSY	328 334 224 216 216 213 181
	172		181 209
RAP2.2		-EENPWELWSLDEIN-FWLEGDE	374

Figure 1. DNA sequence and deduced amino acids of the partial *HvRAF*. Mammalian hemopexin domain signature-like sequence is indicated by solid underline (**A**). Alignment of hemopexin domain signature-like sequences of HvRAF and several other AP2 transcription factors. Identical amino acid residues are shown in black (**B**).

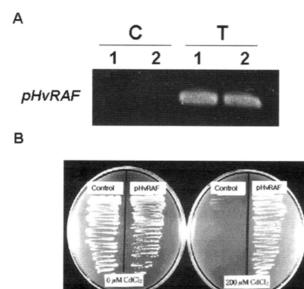


Figure 2. Cadmium tolerance by yeast transformants over-expressing *pHvRAF.* RT-PCR analyses of *pHvRAF* were performed for untransformed yeast *Saccharomyces cerevisiae* strain INVSc1 (C) and yeast transformed with pYES2::*pHvRAF* (T) (**A**). Untransformed yeasts (control) and yeast transformants over-expressing *pHvRAF* (pHvRAF) were grown on SD/-Ura media in absence (left) and presence (right) of 200 μ M CdCl₂ (**B**).

polypeptide of 328 amino acids, containing a putative nuclear localization sequence at the N-terminus, a highly conserved ERF domain, and an acidic (pl = 3.2) C-terminus domain with a novel hemopexin domain signature-like sequence (Jung et al., 2007). Hemopexin is a mammalian serum glycoprotein that binds heme and transports it to the liver (Tolosano and Altruda, 2002). Similar sequences have been found in the C-terminus domain of several AP2 domain-containing proteins, including CBF1-4 (Fig. 1B) (Haake et al., 2002; Jung et al., 2007).

As an initial characterization of the biological functioning of the C-terminus domain in HvRAF, we constructed the pYES2::pHvRAF (partial HvRAF) recombinant plasmid and transformed it into yeast (S. cerevisiae strain INVSc1). This was performed according to the modified lithium acetatemediated method (Gietz et al., 1992). YPD and SD-Ura media were used as the yeast growth substrates. The partial HvRAF contained a C-terminal open reading frame of 113 amino acids (pl = 3.32), including the hemopexin domain signature-like sequence, as well as 361 nucleotides within the 3'-UTR (Fig. 1A). Expression in our yeast transformants was confirmed by RT-PCR analyses (Fig. 2A), using forward (5'-ACACGATGCCGAGGGTCGA-3') and reverse (5'-TCAGAA-AATGGCGCTGTCCA-3') primers. Afterward, we conducted a cadmium-tolerance test of those yeast transformants that indeed harbored pYES2::pHvRAF. In SD-Ura media containing 200 µM CdCl₂, the transformants showed enhanced tolerance to cadmium stress compared with the nontransformants (Fig. 2B). Therefore, we can conclude that the exogenously overexpressed C-terminus region of the HvRAF confers enhanced tolerance to cadmium toxicity in yeast. It remains to be studied how this is accomplished, and whether the same region will confer such tolerance in

higher plants. However, it is tempting to speculate that the hemopexin domain signature-like sequence might be somehow involved in this tolerance mechanism, especially because we have previously demonstrated that 26 amino acids corresponding to this sequence act as a transactivation domain in yeast (Jung et al., 2007).

ACKNOWLEDGMENTS

This study was supported by a research grant (CG1520) from the Crop Functional Genomics Center of the 21st Century Frontier Research Program funded by the Korean Ministry of Science and Technology. Jinwook Jung was supported by the Korean Ministry of Education through the Brain Korea 21 Project.

Received March 5, 2007; accepted March 26, 2007.

LITERATURE CITED

- Chen W, Provart NJ, Glazebrook J, Katagiri F, Chang HS, Eulgem T, Mauch F, Luan S, Zou G, Whitham SA, Budworth PR, Tao Y, Xie Z, Chen X, Lam S, Kreps JA, Harper JF, Si-Ammiur A, Mauch-Mani B, Heinlein M, Kobayashi K, Hohn T, Dangl JL, Wang X, Zhu T (2002) Expression profile matrix of *Arabidopsis* transcription factor genes suggests their putative functions in response to environmental stresses. Plant Cell 14: 559-574
- Clemens S, Kim EJ, Neumann D, Schroeder JI (1999) Tolerance to toxic metals by a gene family of phytochelatin synthases from plants and yeast. EMBO J 18: 3325-3333
- Clijstersw H, van Assche F (1985) Inhibition of photosynthesis by metals. Photosynth Res 7: 31-40
- Cobbett CS, May MJ, Howden R, Rolls B (1998) The glutathionedeficient, cadmium-sensitive mutant, cad2-1, of *Arabidopsis thaliana* is deficient in γ-glutamylcysteine synthase. Plant J 16: 73-78
- Gietz D, St Jean A, Woods RA, Schiestl RH (1992) Improved method for high efficiency transformation of intact yeast cells. Nucl Acids Res 20: 1425
- Ha SB, Smith AP, Howden R, Dietrich WM, Bugg S, O'Connell MJ, Goldsbrough PB, Cobbett CS (1999) Phytochelatin synthase genes from *Arabidopsis* and the yeast, *Schizosaccharomyces pombe*. Plant Cell 11: 1153-1164
- Haake V, Cook D, Riechmann JL, Pineda O, Thomashow MF, Zhang JZ (2002) Transcription factor CBF4 is a regulator of drought adaptation in *Arabidopsis*. Plant Physiol 130: 639-648
- Jung J, Won SY, Suh SC, Kim H, Wing R, Jeong Y, Hwang I, Kim M (2007) The barley ERF-type transcription factor, HvRAF, confers enhanced pathogen resistance and salt tolerance in Arabidopsis. Planta 225: 575-588
- Keck RW (1978) Cadmium alteration of root physiology and potassium ion fluxes. Plant Physiol 116: 1413-1420
- Kim YJ, Chang KS, Lee MR, Kim JH, Lee CE, Jeon YJ, Choi JS, Shin HS, Hwang S (2005) Expression of tobacco cDNA encoding phytochelatin synthase promotes tolerance to and accumulation of Cd and As in Saccharomyces cerevisiae. J Plant Biol 48: 440-447
- Krupa Z, Oquist G, Humur NPA (1993) The effects of cadmium on photosynthesis of *Phaseolus vulgaris* L.: A fluorescence analysis. Physiol Plant 88: 626-630
- Lee J, Shim D, Song WY, Hwang I, Lee Y (2004) Arabidopsis metal-

lothioneins 2a and 3 enhance resistance to cadmium when expressed in *Vicia faba* guard cell. Plant Mol Biol **54**: 805-815

- Li ZS, Szczypka M, Lu YP, Thiele DJ, Rea PA (1996) The yeast cadmium factor protein (YCF1) is a vacuolar glutathione S-conjugate pump. J Biol Chem 271: 6509-6517
- Lu YP, Li ZS, Rea PA (1997) AtMRP1 gene of *Arabidopsis* encodes a glutathione S-conjugate pump: Isolation and functional definition of a plant ATP-binding cassette transporter gene. Proc Natl Acad Sci USA 94: 8243-8248
- Somashekaraiah BV, Padmaja K, Prasad ARK (1992) Phytotoxicity of cadmium ions on germination of seedlings of mung bean (*Phaseolus vulgaris*): Involvement of lipid peroxides in chloro-

phyll degradation. Physiol Plant 85: 85-89

- Song WY, Sohn EJ, Martinoia E, Lee YJ, Yang YY, Jasinski M, Cyrille F, Hwang I, Lee Y (2003) Engineering tolerance and accumulation of lead and cadmium in transgenic plants. Nat Biotech 21: 914-919
- Tolosano E, Altruda F (2002) Hemopexin: Structure, function, and regulation. DNA Cell Biol 21: 297-306
- Vatamaniuk OK, Mari S, Lu YP, Rea PA (1999) AtPCS1, a phytochelatin synthase from *Arabidopsis*: Isolation and in vitro reconstitution. Proc Natl Acad Sci USA 96: 7110-7115
- Zenk MH (1996) Heavy metal detoxification in higher plants. Gene 179: 21-30